

SUPPLEMENTARY MATERIAL

**Micro-Raman Spectroscopy Reveals the Presence of Octacalcium Phosphate and  
Whitlockite in Association with Bacteria-Free Zones Within the Mineralized Dental  
Biofilm**

**Brief title:**

**Revisiting the Mineralized Dental Biofilm**

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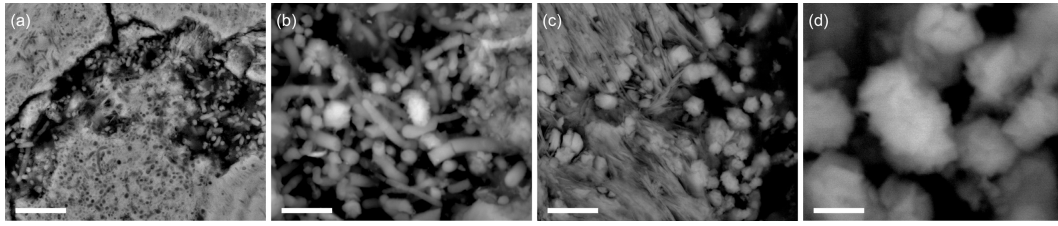
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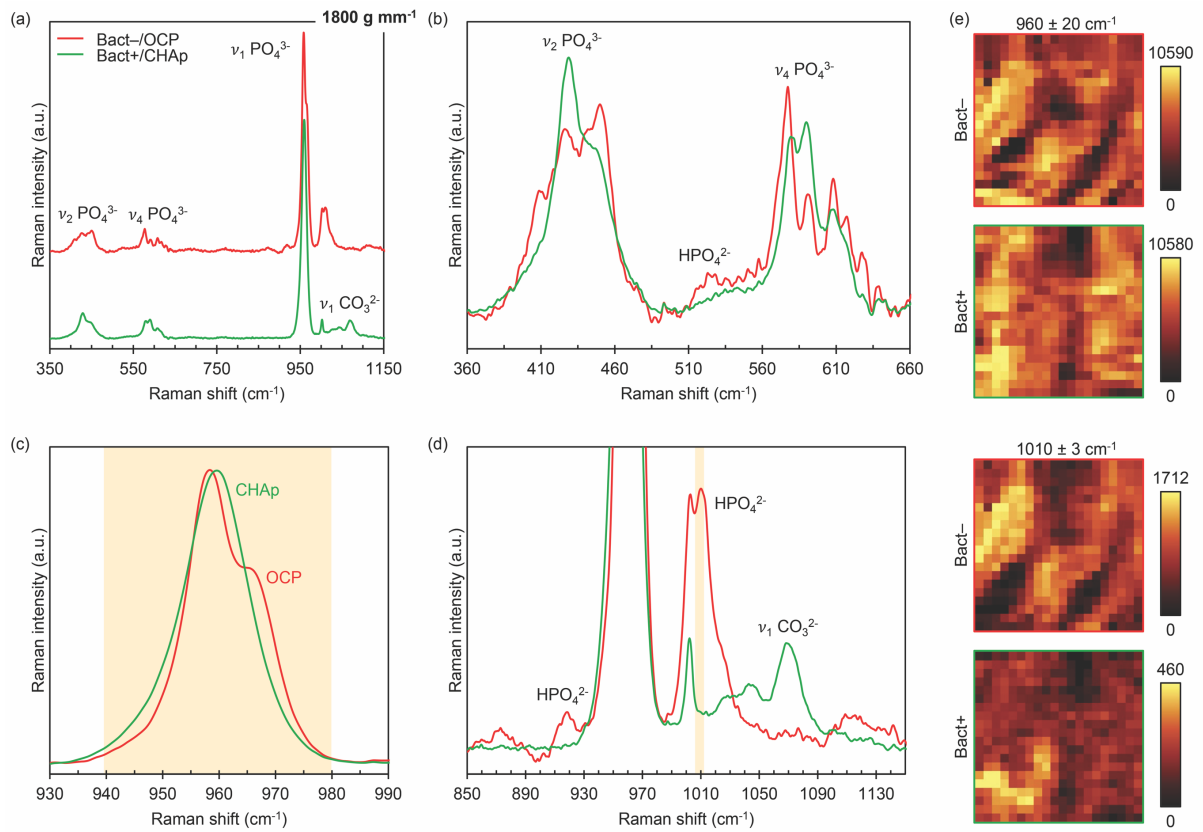
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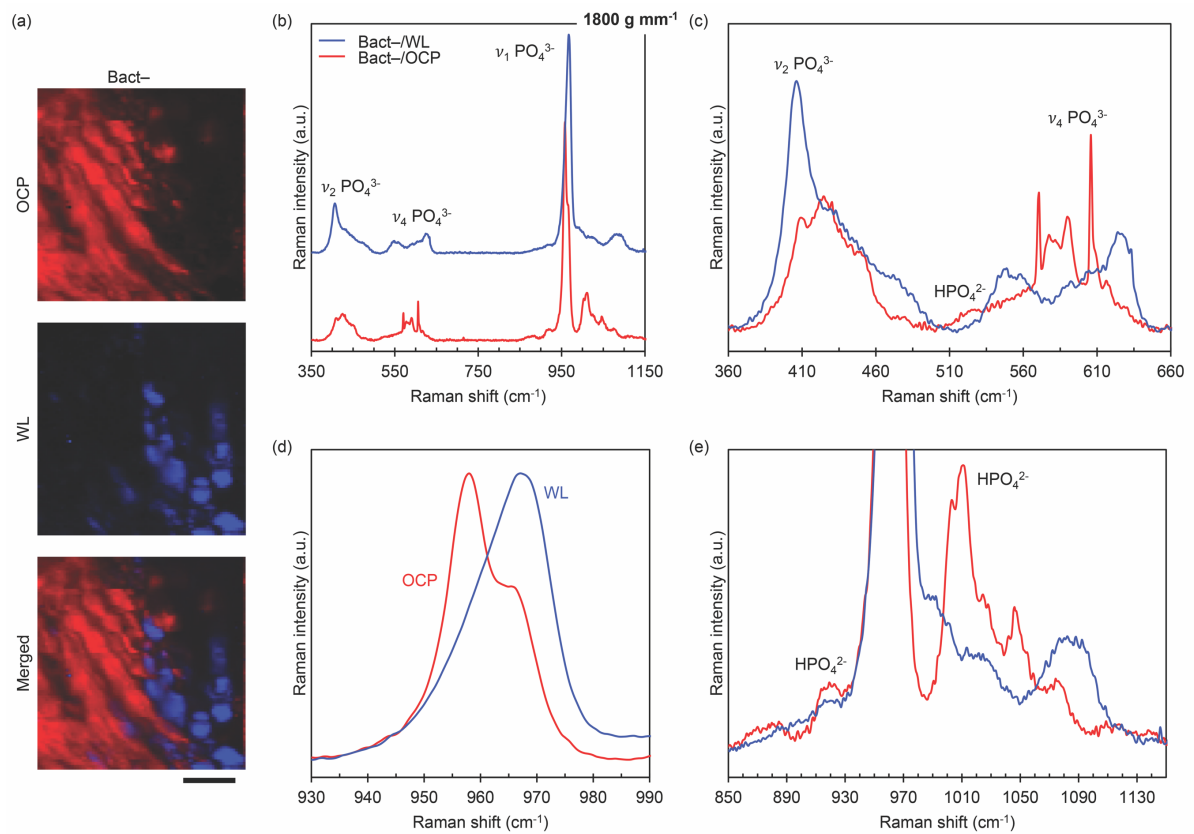
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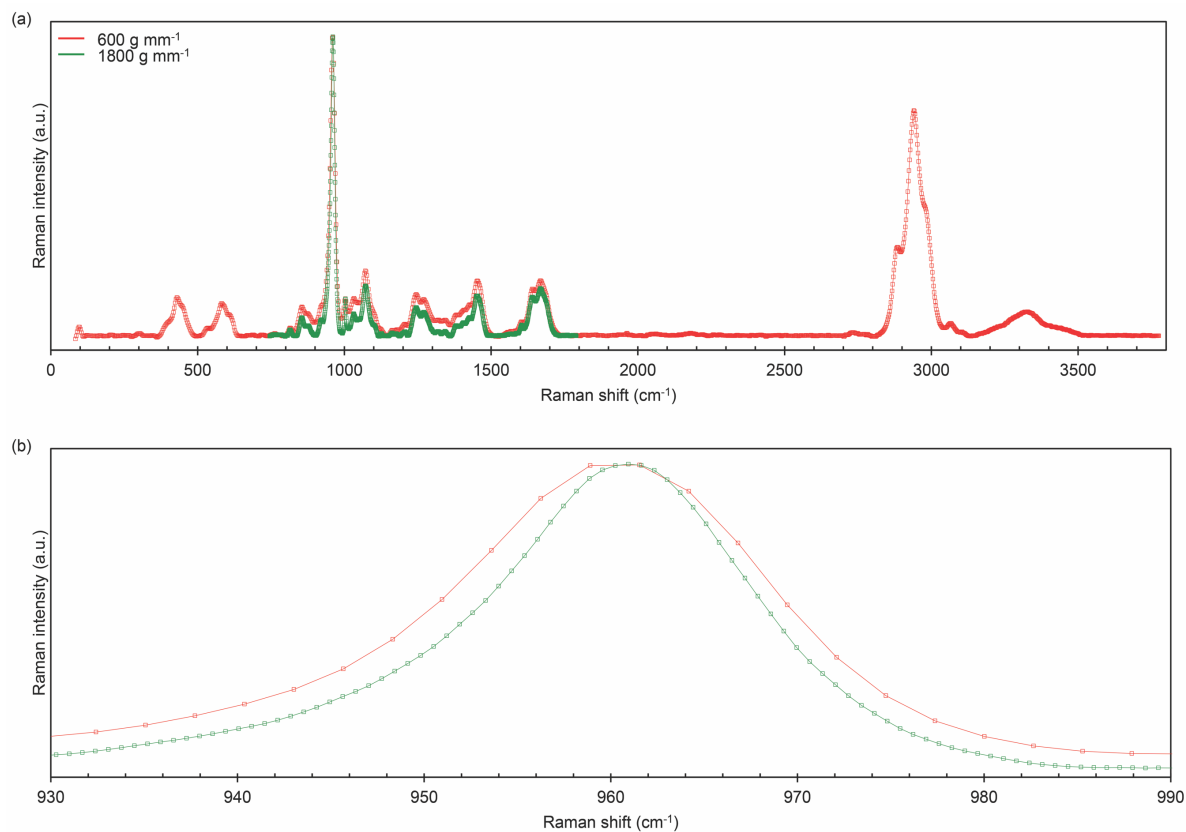
**Fig. S1: Backscattered electron scanning electron microscopy.** (a) Bacteria-containing zone (Bact+) showing mineralized bacteria and partial extramicrobial mineralization. (b) Intramicrobial mineralization is seen as smooth, solid mineral deposits resembling the morphologies of bacteria found in dental plaque. (c) Bacteria-free zone (Bact-) showing mineral platelets and micrometer-sized mineral nodules. (d) The mineral nodules appear considerably less smooth than mineralized bacteria. Scale bars in a = 10  $\mu\text{m}$ ; b, c = 5  $\mu\text{m}$ ; and d = 1  $\mu\text{m}$ .



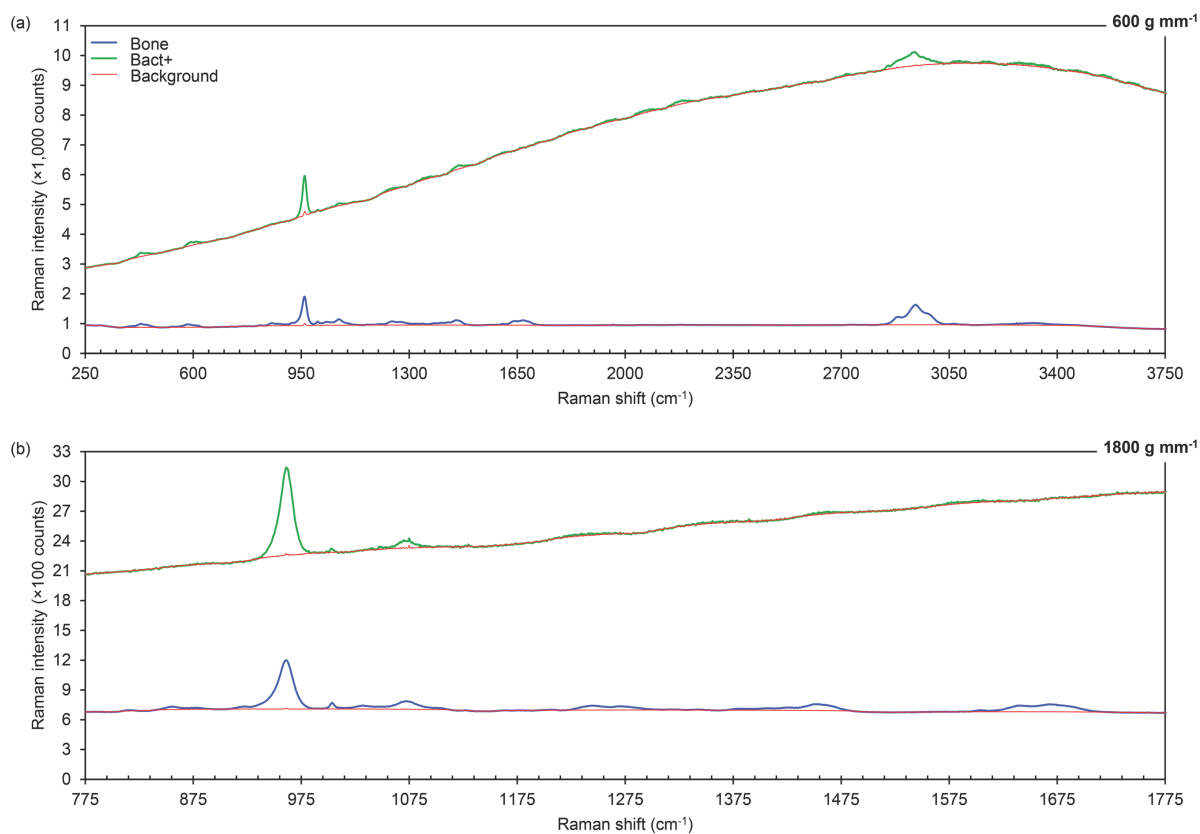
**Fig. S2: Raman spectroscopy.** (a) Average spectra of Bact+ and Bact- (1,800 g mm<sup>-1</sup> grating, 317–1,423 cm<sup>-1</sup> spectral range, spectral center at 900 cm<sup>-1</sup>, 10 s integration time per pixel). Bact+ are comprised of carbonated apatite (CHAp). Bact- are predominantly made up of octacalcium phosphate (OCP). 5 μm × 5 μm ROI, 400 spectra; (*n* = 2). (b) Detail of the 360–660 cm<sup>-1</sup> region ( $\nu_2$  PO<sub>4</sub><sup>3-</sup> and  $\nu_4$  PO<sub>4</sub><sup>3-</sup>). (c) Detail of the 930–990 cm<sup>-1</sup> region ( $\nu_1$  PO<sub>4</sub><sup>3-</sup>; normalized intensities). (d) Detail of the 850–1,150 cm<sup>-1</sup> region (HPO<sub>4</sub><sup>2-</sup>,  $\nu_1$  CO<sub>3</sub><sup>2-</sup>). (e) The example shows Raman maps of  $\nu_1$  PO<sub>4</sub><sup>3-</sup> (960 ± 20 cm<sup>-1</sup>) and HPO<sub>4</sub><sup>2-</sup> (1,010 ± 3 cm<sup>-1</sup>) in Bact+ and Bact-. Bact+ exhibit very low HPO<sub>4</sub><sup>2-</sup> signal, indicating the absence of OCP. At Bact-, HPO<sub>4</sub><sup>2-</sup> signal is significantly higher (vs. Bact+) and the spatial distribution closely follows that of  $\nu_1$  PO<sub>4</sub><sup>3-</sup>. The color scale bars show charge-coupled device (CCD) detector counts.



**Fig. S3: Raman spectroscopy.** (a) Raman maps of octacalcium phosphate (OCP), whitlockite (WL), and color-merge of OCP (red) and WL (blue) in Bact-. 20 μm × 20 μm ROI, 6,400 spectra. (b) OCP and WL spectra corresponding to the Raman maps (shown in a). (c) Detail of the 360–660 cm<sup>-1</sup> region ( $\nu_2$  PO<sub>4</sub><sup>3-</sup> and  $\nu_4$  PO<sub>4</sub><sup>3-</sup>). (d) Detail of the 930–990 cm<sup>-1</sup> region ( $\nu_1$  PO<sub>4</sub><sup>3-</sup>; normalized intensities). (e) Detail of the 850–1,150 cm<sup>-1</sup> region (HPO<sub>4</sub><sup>2-</sup>,  $\nu_1$  CO<sub>3</sub><sup>2-</sup>). Scale bar in a = 5 μm.



**Fig. S4: Comparison of spectral range and spectral resolution achieved with 600 g mm<sup>-1</sup> and 1,800 g mm<sup>-1</sup> gratings.** (a) The example shows average spectra of native bone. The spectral range of the 600 g mm<sup>-1</sup> (84–3,775 cm<sup>-1</sup> spectral range; spectral center at 2,150 cm<sup>-1</sup>) is significantly broader than the 1,800 g mm<sup>-1</sup> grating (745–1,790 cm<sup>-1</sup> spectral range; spectral center at 1,250 cm<sup>-1</sup>). (b) Due to a smaller step-size, high resolution spectra allow detection of fine features, e.g., peak broadening, peak shifts, and band splitting, such as of the  $\nu_1$  PO<sub>4</sub><sup>3-</sup> peak (here, centered around 960 cm<sup>-1</sup>).



**Fig. S5: Background fluorescence emission by Bact+ and native bone.** (a–b) Bact+ generate significantly higher background fluorescence than native bone with both 600 g mm<sup>-1</sup> (a) and 1,800 g mm<sup>-1</sup> (b) gratings. Compared to native bone, the background fluorescence emission from Bact+ increases dramatically at higher wavenumbers.