**Appendix: Supplemental Methods**

*Quantitative real-time PCR.* Swabs were collected and stored immediately in Buffer AVL at -80°C prior to RNA extraction and PCR testing. RNA was extracted using QIAmp® Viral RNA Mini Kits (QIAGEN, Hilden, Germany) according to manufacturer’s instructions. 20µL PCR reactions were set up containing 0.5µL (20µM) each of SARS-CoV-2 N1 forward and reverse primer, 0.5µL (5µM) SARS-CoV-2 N1 probe, 5µL TaqManTM Fast Virus 1-Step Master Mix (ThermoFisher Scientific, Waltham, MA), 8.5µL molecular grade water, and 5µL RNA template. PCR was conducted in technical triplicates in 96-well plates using Applied BiosystemsTM QuantStudioTM 5 Real-Time PCR System (ThermoFisher Scientific, Waltham, MA). Reverse transcription was performed at 50°C for 5 minutes and initial denaturation at 95°C for 20 seconds, followed by 40 cycles of denaturation at 95°C for 3 seconds and annealing/extension at 60°C for 30 seconds.