**Supplemental Material. Pulsed field gel electrophoresis (PFGE)**

MRSAisolates were grown in brain heart infusion broth overnight in a 35°C incubator and the resultant suspension adjusted to an OD590 of 45–55% transmittance. 7 ml of the cell suspension was centrifuged for 10 min at 2,000 rpm and the supernatant discarded. Cells were re-suspended in 500 µl of EET buffer (100 mM EDTA, 10 mM EGTA, 10 mM tris, pH 8.0), vortexed and allowed to acclimate in a 67°C water bath. 500 µl of 1.6% low-melt sample agarose (SeaPlaque® GTG Agarose, Lonza, Basel, Switzerland) was added to the cell suspension and gently mixed. Two sample plugs were prepared within a PFGE insert (BioRad, Hercules, CA) for each isolate and allowed to solidify at room temperature.

Cells were lysed within the sample plugs in EET-LL solution (100 mM EDTA, 10 mM EGTA, 10 mM tris pH 8.0 with 50 mg/µl lysozyme and 25 units/mL lysostaphin) for 4 hours at 30°C. The lysis buffer was removed and lysis terminated by addition of 1 ml of EET-SP solution (100 mM EDTA, 10 mM EGTA, 10 mM Tris pH 8.0 with 1% sodium dodecyl sulfate and 1 mg/ml proteinase K) and subsequent incubation at 50°C for 18 hours. EET-SP was removed and the sample plugs were washed four times with 40 ml of TE wash buffer containing 10 mM tris 1mM EDTA pH 8.0.

Approximately 1/5th of each insert was transferred to a 1.5 ml microcentrifuge tube containing a 1X solution of restriction buffer J and 0.3 units/µl of SmaI restriction enzyme (Roche Applied Science, Indianapolis, IN) and incubated at 25°C for 4 hours. Following incubation, the enzyme mixture was removed and 1 ml of a 0.5X electrophoresis running buffer (TAE - 200 mM Tris 100 mM acetic acid 0.5 mM EDTA) was added to each restricted plug and incubated for 10 min at room temperature. Following incubation, TAE was removed and inserts placed into a 67°C water bath to melt. Up to 40 µl of each insert were pipetted into wells of a 21 X 14 cm 1% agarose (SeaKem® HGT Agarose, Lonza) analytical gel. Two liters of TAE were added to the electrophoresis chamber and chilled to 14°C. The analytical gel was loaded into the electrophoresis chamber and the CHEF DRII apparatus run for 15 hours at 6 v/cm, 120° angle with initial and final switch times of 10 and 50 sec, respectively. Following electrophoresis, the gel was stained in ethidium bromide and rinsed with deionized water. The gel image was captured on a GelDoc XR system (BioRad). Comparison of PFGE patterns was performed manually and representative isolates from suspected outbreak clusters were tested on serial gels. Isolates were classified as indistinguishable if there were no differences noted, and different if differences were noted.