

## Supplemental Content

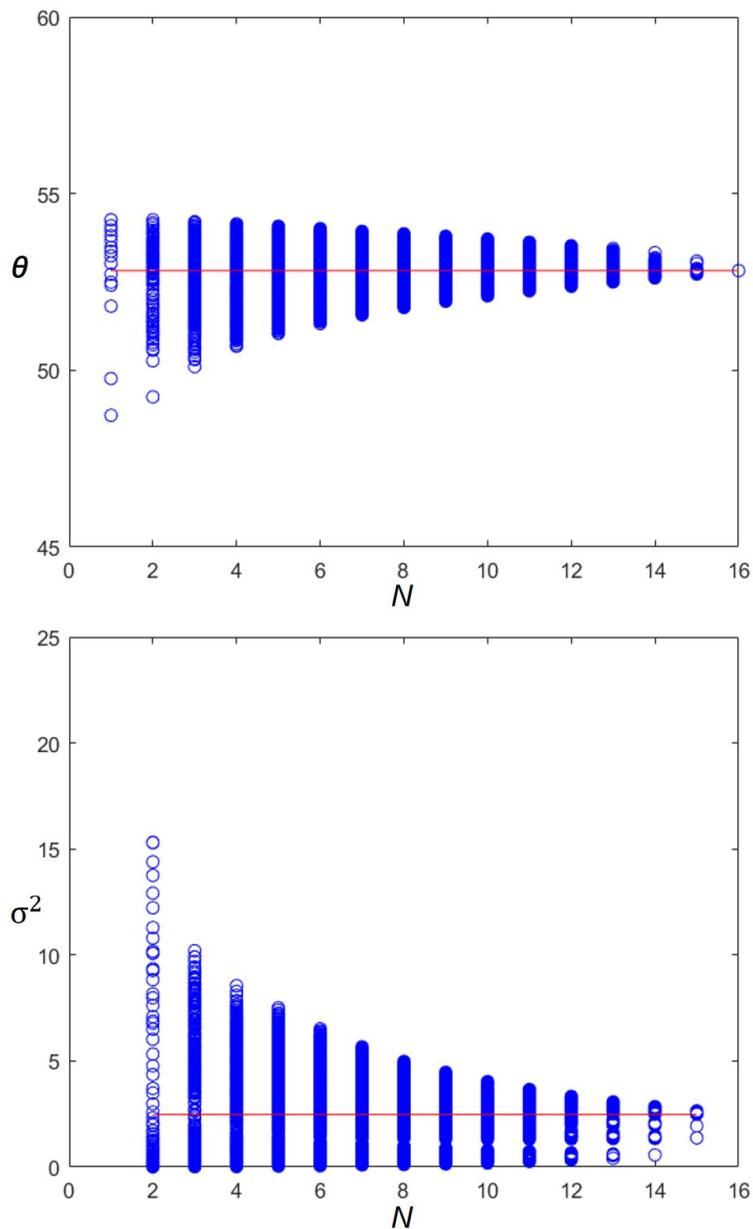


Figure 1: Sample size justification. Rarefaction to assess the effect of family richness on our estimation of variance: Variance  $\sigma^2$  and  $\theta$  values calculated for different numbers of sampled moth families ( $N$ ), with a random sample of  $N$  families at each level of rarefaction. Convergence of values can be seen with increased sampling effort.

*Moth-inspired particle capture*

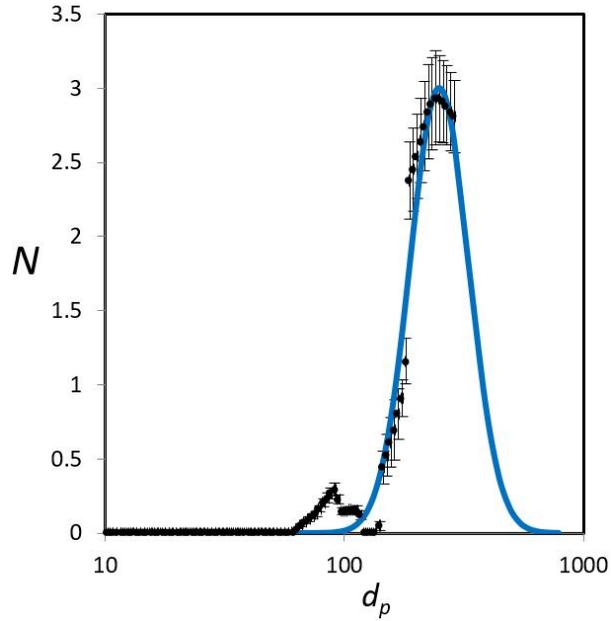


Figure 2: Number distribution  $N$  of particles in millions per  $\text{cm}^3$  used in experiment by particle diameter in nanometers  $d_p$  with log-normal trend in blue.

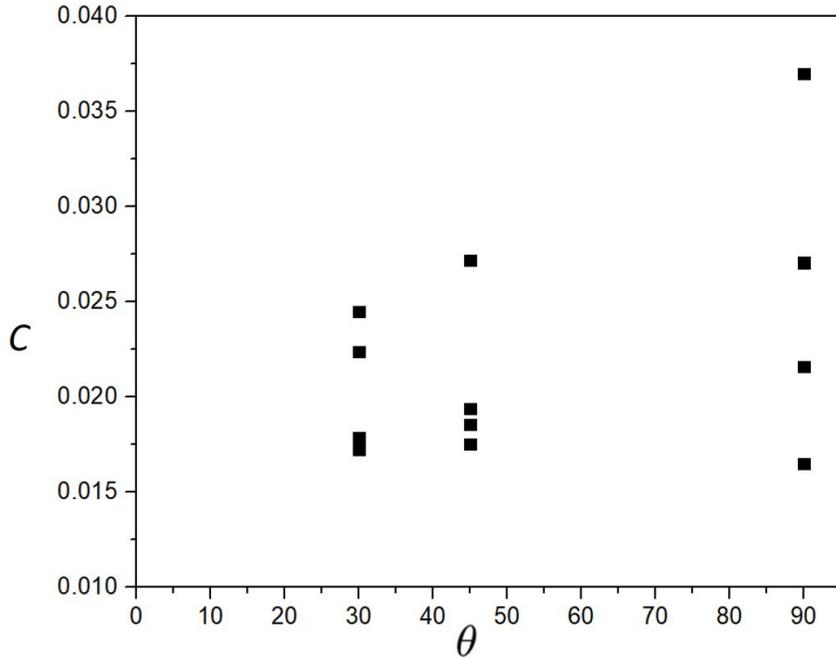


Figure 3: PH strips experimental data: Concentration of hydrogen ions at different cylinder angles. Concentration  $C$  is in mol H+ per liter air. Angle of cylinder with respect to bulk flow direction  $\theta$  is in degrees

Table 1: Variable List

Symbol	Name	Description
$A$	Projected area	Frontal area of cylinder projected onto y-z plane, in $\mu\text{m}^2$
$A_s$	Surface area	Estimated cylinder surface area ignoring end effects
$C$	Concentration	$3 \times 10^9$ pheromone molecules per $\text{m}^3$ to elicit moth response
$d$	Diameter	Moth stalk diameter of $136 \pm 36 \mu\text{m}$ , branch diameter of $43 \pm 14 \mu\text{m}$ , and sensilla diameter of $5.5 \pm 1.4 \mu\text{m}$ .
$D$	Diffusion coefficients	$5.74 \times 10^{-10} \text{ m}^2/\text{s}$ for humid air in wind tunnel experiment $2.5 \times 10^{-5} \text{ m}^2/\text{s}$ to $2.5 \times 10^{-6} \text{ m}^2/\text{s}$ for odor and pheromone
$d_c$	Collector Diameter	$135 \mu\text{m}$ diameter cylinder used as mimic in wind tunnel experiment
$d_p$	Particle Diameter	Diameter of drops used in wind tunnel experiment: $2*r_p$
$L$	Length of cylinder	Fiber length of 3 mm used in wind tunnel experiment
$L_b$	Branch length	Average length of antennae branch: $750 \pm 395 \mu\text{m}$
$L_s$	Sensilla length	Average length of antennae sensilla: $70 \pm 25 \mu\text{m}$
$L_{st}$	Stalk length	Average length of antennae stalk: $9 \pm 2 \text{ mm}$
$M$	Mass of moth	Mass of dried moth in grams
$N$	Number of molecules	Estimated number of pheromone molecules needed to excite
$N_b$	Total number of branches	Average number of branches per antennae stalk: $85 \pm 20$
$N_s$	Total number of sensilla	Average number of sensilla per antennae branch: $28 \pm 14$
$r_p$	Drop radius	Radius of drops used in wind tunnel experiment: $5 \mu\text{m}$
$S_b$	Branch spacing	Center to center distance between branches: $116 \pm 35 \mu\text{m}$
$S_s$	Sensilla spacing	Center to center distance between sensilla: $24 \pm 4.4 \mu\text{m}$
$t$	Travel time	Average time a particle travels next to the collection cylinder
$T$	Detection time	Estimated time to collect required number of molecules
$U$	Air velocity	$0.5 \text{ m/s}$ in wind tunnel experiment. $0.75\text{-}1 \text{ m/s}$ moth flight
$x$	Diffusion distance	Distance that a particle can travel by diffusion in a given time
$\eta$	Capture efficiency	Single cylindrical collector capture efficiency: 2-2.6
$\nu$	Kinematic viscosity	Air kinematic viscosity at room temperature: $1.46 \times 10^{-5} \text{ m}^2/\text{s}$
$\rho$	Density	Air density at room temperature. $1.23 \text{ kg/m}^3$
$\phi$	Stalk angle	Angle between stalk and flight direction in x-z plane in degrees
$\theta$	First branch angle	Angle between branch and stalk in x-z plane in degrees
$\gamma$	Second branch angle	Angle between branch and stalk in x-y plane in degrees
$\alpha$	Lingering coefficient	Dimensionless term to account for the deflection of the streamlines that encourages flow to linger around the cylinder
$\kappa$	Percent covered	Percentage of fiber covered per second

Variables and associated values

*Moth-inspired particle capture*

Table 2: Moth Specimen Information

	<b>Identification</b>	<b>Family</b>	<b>Location</b>	<b>Year</b>
1	Unidentified Blastobasidae sp. 1	Blastobasidae	Heredia, Costa Rica	-
2	Unidentified Blastobasidae sp. 2	Blastobasidae	Heredia, Costa Rica	2003
3	<i>Apatelodes sp.</i>	Bombycidae	Heredia, Costa Rica	1999
4	<i>Apatelodes torrefacta</i>	Bombycidae	Nantahala region, NC	2015
5	<i>Zeuzera sp.</i>	Cossidae	-	-
6	<i>Inguromorpha itzalana</i>	Cossidae	Cochise Co, AZ	2010
7	<i>Symphlebia pyrgion</i>	Erebidae	Heredia, Costa Rica	2005
8	<i>Lymantria dispar</i>	Erebidae	Massachusetts lab	2015
9	<i>Halysidota tessellaris</i>	Erebidae	Nantahala region, NC	2015
10	<i>Apantesis sp.</i>	Erebidae	Nantahala region, NC	2015
11	<i>Leucanopsis sp. 1</i>	Erebidae	Heredia, Costa Rica	2001
12	<i>Hormisa orciferalis</i>	Erebidae	Windham Co, CT	2011
13	Unidentified Erebidae sp.	Erebidae	Heredia, Costa Rica	2007
14	<i>Dasychira pinicola</i>	Erebidae	Bamstable MA	1995
15	Unidentified Geometridae sp. 1	Geometridae	Nantahala region, NC	2015
16	<i>Lytrosis unitaria</i>	Geometridae	Nantahala region, NC	2015
17	<i>Epimecis hortaria</i>	Geometridae	Nantahala region, NC	2015
18	<i>Lytrosis unitaria</i>	Geometridae	Nantahala region, NC	2015
19	Unidentified Geometridae sp. 2	Geometridae	Nantahala region, NC	2015
20	Unidentified Geometridae sp. 3	Geometridae	Nantahala region, NC	2015
21	Unidentified Geometridae sp. 3	Geometridae	Nantahala region, NC	2015
22	Unidentified Geometridae sp. 2	Geometridae	Nantahala region, NC	2015
23	Unidentified Geometridae sp. 3	Geometridae	Nantahala region, NC	2015
24	Unidentified Geometridae sp. 4	Geometridae	Nantahala region, NC	2015
25	<i>Lytrosis unitaria</i>	Geometridae	Nantahala region, NC	2015
26	Unidentified Geometridae sp. 4	Geometridae	Nantahala region, NC	2015
27	Unidentified Geometridae sp. 5	Geometridae	Nantahala region, NC	2015
28	<i>Itame pustularia</i>	Geometridae	Nantahala region, NC	2015
29	<i>Itame pustularia</i>	Geometridae	Nantahala region, NC	2015
30	<i>Itame pustularia</i>	Geometridae	Nantahala region, NC	2015
31	<i>Anticlea sp.</i>	Geometridae	Heredia, Costa Rica	2000
32	<i>Glena cribrataria</i>	Geometridae	Dukes Co., MA	1998
33	<i>Caripeta divisata</i>	Geometridae	Litchfield Co., CT	1998
34	<i>Aepytus sp. 1</i>	Hepialidae	Heredia, Costa Rica	2004
35	<i>Malacosoma americana</i>	Lasiocampidae	Nantahala region, NC	2015
36	<i>Euglyphis barda</i>	Lasiocampidae	Heredia, Costa Rica	2003
37	<i>Euclea delphinii</i>	Limacodidae	New London Co, CT	2012
38	<i>Lagoa crispata</i>	Megalopygidae	Windham Co, CT	2008
39	<i>Norape tenera</i>	Megalopygidae	Cochise Co, AZ	1999
40	<i>Lirimiris lignitecta</i>	Notodontidae	-	-
41	<i>Thyridopteryx ephemeraeformis</i>	Psychidae	Ohio	2007
42	<i>Rothschildia sp.1</i>	Saturniidae	Heredia, Costa Rica	2001
43	<i>Automeris sp.</i>	Saturniidae	Nantahala region, NC	2015
44	<i>Asthenia transversaria</i>	Saturniidae	Heredia, Costa Rica	2004
45	<i>Perigonia pittieri</i>	Sphingidae	Heredia, Costa Rica	2002

Collection information for 45 unique moth specimens spanning 15 families. Information marked with ”-” was not supplied when specimen was received. Four additional moths were measured but unidentified.

Table 3: Moth Antennae Dimensions

<b>Dimension</b>	<b>Equation</b>
Stalk Length	$L_{st}=19,300M^{0.20}$
Branch Length	$L_b=2,070M^{0.28}$
Sensilla Length	$L_s=104M^{0.079}$
Stalk Diameter	$d_{st}=157M^{0.029}$
Branch Diameter	$d_b=63.8M^{0.11}$
Sensilla Diameter	$d_s=5.96M^{0.023}$
Branch Spacing	$S_b=167M^{0.051}$
Sensilla Spacing	$S_s=20.7M^{-0.052}$

Allometric equations of moth antennae dimensions presented in Fig. 2. All dimensions are in micrometers.  $M$  represents dry moth mass in grams.

Table 4: Increased path length values

$\theta$	$w^*$
15	2.00
30	0.69
45	0.28
60	0.12
75	0.06
90	0.00

Dimensionless increase in path length  $w^*$  particles must travel around a cylinder held at angle  $\theta$  into the wind.