

Occurrence of methicillin-resistant *Staphylococcus aureus* (MRSA) in “coalho” cheeses produced in Brazil

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SUPPLEMENTARY FILE

Material & Methods

Isolation of Staphylococcus aureus

In the laboratory, with the aid of sterile scalpels, the samples were cut and weighed. 25 g of random portions of the food were collected, including surface and depth. Subsequently, 225 ml of 0.1% peptone salt solution were added, obtaining a 1:10 dilution; the samples were subjected to the process of homogenization in the “stomacher” for 60 seconds, and from this dilution, serial dilutions up to 1×10^{-5} were performed.

After all dilutions were prepared, a 0.1 ml aliquot of each was transferred to the surface of a petri dish containing Baird-Parker agar (LABORCLIN, BRAZIL), enriched with egg yolk emulsion and potassium tellurite, and spread with the aid of a Drigalski loop, always in duplicate.

All inoculated plates were taken to the bacteriological incubator and incubated for 48 hours at 35 °C - 37 °C. After the incubation period, the plates that had between 20 and 200 colonies were selected and the colonies were categorized, being divided into typical and atypical. From each plate, which the two types of colonies existed, 3 typical and 2 atypical colonies were removed with the aid of a sterile platinum handle; from the plates which there were only typical colonies, 5 colonies were removed; and the plates which only atypical colonies were observed, there was no removal of colonies for molecular tests, only for counting the CFU/g of the sample.

Multiplex PCR

For the reaction, 30 ng of genomic DNA were used, 20 pmol of specific primers for each gene, 160 µg of each dNTP, 3 mM magnesium chloride (MgCl₂), 50 mM potassium chloride (KCl), 10 mM Tris-HCl (pH 9.0), and 1.2U of Taq DNA Polymerase (Invitrogen, Brazil), generating a reaction product with a total volume of 25 µl.

The amplification reactions took place in a thermocycler (Byocycler) programmed for 30 complete thermal cycles; and the *S. aureus* strains standard ATCC 43300, ATCC 13565, ATCC 14458, ATCC 19095 and ATCC 13566, possessing the *mecA* and *nuc* genes, were used as positive controls of *sea*, *seb*, *sec*, and *tst1*, respectively.

After the reactions, the amplification products (amplicons) were analyzed by electrophoresis in 1.5% (w/v) agarose gel, marked with ethidium bromide (1.5 mg/ml), and observed in a UV transilluminator. The primers used in the reactions are described in Table S1.

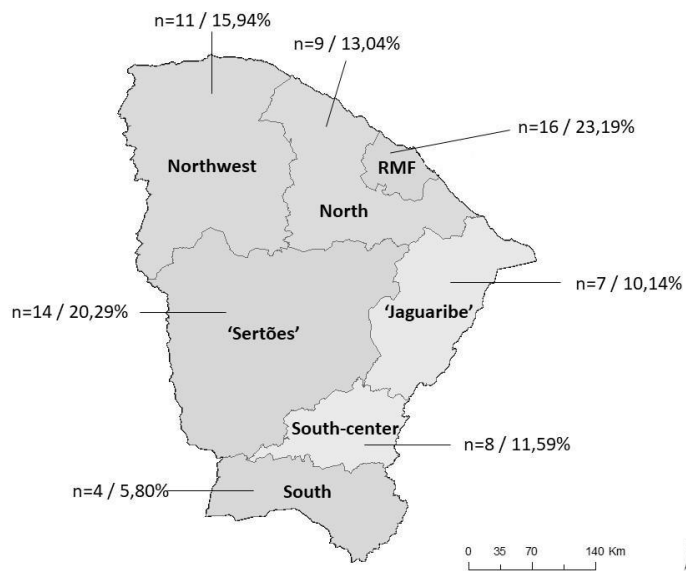
Table S1:

Sequence of primers used and sizes of amplicons in number of base pairs.

Gene	Primer	Sequence 5' – 3'	Amplicon (pb)	Reference
<i>nuc</i>	<i>nuc</i> -F	GCGATTGATGGTGATACGGTT	279	KATEETE <i>et al.</i> , 2010
	<i>nuc</i> -R	AGCCAAGCCTTGACGAACTAAAGC		
<i>mecA</i>	<i>MecA</i> -F	GTAGAAATGACTGAACGTCCGATAA	310	MCCLURE <i>et al.</i> , 2006
	<i>MecA</i> -R	CCAATTCCACATTGTTTCGGTCTAA		
<i>blaZ</i>	<i>blaZ</i> -F	AAGAGATTTGCCTATGCTTC	517	SAWANT; GILLESPIE; OLIVER, 2009
	<i>blaZ</i> -R	GCTTGACCACTTTTATCAGC		
<i>sea</i>	<i>sea</i> -F	TAAGGAGGTGGTGCCTATGG	180	CREMONESI <i>et al.</i> , 2005
	<i>sea</i> -R	CATCGAAACCAGCCAAAGTT		
<i>seb</i>	<i>seb</i> -F	CATCGAAACCAGCCAAAGTT	478	JARRAUD <i>et al.</i> , 2002
	<i>seb</i> -R	TCGCATCAAACCTGACAAACG		
<i>sec</i>	<i>sec</i> -F	ACCAGACCCTATGCCAGATG	371	CREMONESI <i>et al.</i> , 2005
	<i>sec</i> -R	TCCCATTATCAAAGTGTTTCC		
<i>tst1</i>	<i>tst1</i> -F	ATGGCAGCATCAGCTTGATA	300	AKINEDEN <i>et al.</i> 2001
	<i>tst1</i> -R	TTTCCAATAACCACCCGTTT		

Figure S1:

Map of Ceará with the distribution of the number of *S. aureus* strains in ‘coalho’ cheese samples and the percentage by mesoregion of the state of Ceará - Brazil.



Adapted from Map of Mesoregions of Ceará, available at: <http://www.baixarmapas.com.br/mapa-de-mesorreioes-do-ceara/>. Accessed on July 27, 2020

References

- Akineden O, Annemüller C, Hassan AA, Lämmle C, Wolter W & Zschöck M 2001 Toxin genes and other characteristics of *Staphylococcus aureus* isolates from milk of cows with mastitis. *Clinical and Diagnostic Laboratory Immunology* **8** 959-964
- Cremonesi P, Luzzana M, Brasca M, Morandi S, Lodi R, Vimercati C, Agnellini D, Caramenti G, Moroni P & Castiglioni B 2005 Development of a multiplex PCR assay for the identification of *Staphylococcus aureus* enterotoxigenic strains isolated from milk and dairy products. *Molecular and Cellular Probes* **19** 299–305
- Jarraud S, Mougel C, Thioulouse J, Lina G, Meugnier H, Forey F, Nesme X, Etienne J & Vandenesch F 2002 Relationships between *Staphylococcus aureus* genetic background, virulence factors, *agr* groups (alleles), and human disease. *Infection and Immunity* **70** 631–641
- Kateete DP, Kimani CN, Katabazi FA, Okeng A, Okee MS, Nanteza A, Joloba ML & Najjuka FC 2010 Identification of *Staphylococcus aureus*: DNase and Mannitol salt agar improve the efficiency of the tube coagulase test. *Annals of Clinical Microbiology and Antimicrobials* **9**
- McClure JA, Conly JM, Lau V, Elsayed S, Louie T, Hutchins W & Zhang K 2006 Novel multiplex PCR assay for detection of the staphylococcal virulence marker Panton-Valentine leukocidin genes and simultaneous discrimination of methicillin-susceptible from resistant staphylococci. *Journal of Clinical Microbiology* **44** 1141-1144
- Sawant AA, Gillespie BE & Oliver SP 2009 Antimicrobial susceptibility of coagulase-negative *Staphylococcus* species isolated from bovine milk. *Veterinary Microbiology* **134** 73-81

