| he production of dulce de leche |
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| reduction in dulce de leche |
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27 Introduction

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Products with low sugar content are justified by recent studies correlating the decrease in 29 dietary sugar (mainly sucrose) with the prevention, combat and recovery of a series of 30 pathologies such as diabetes, hypertension, obesity, cardiovascular diseases, metabolic 31 syndrome, dementia, depression, anxiety, cancer, dental problems, ulcers, hiatal hernia, 32 Crohn's disease, irritable bowel syndrome, dermatitis, liver disease (Chow, 2017; Della 33 Corte et al., 2018; Khan & Sievenpiper, 2016; Lenne & Mann, 2020). These products 34 with low content or without sugar meet the demand for healthy foods, however they can 35 36 present different functional, rheological, sweetness, stability, particle size distribution and 37 texture when compared to traditional products, thus needing to use other compounds that provide body, texture and sweetness to it (Belščak-Cvitanović et al., 2015). 38

In order to avoid the problems associated with excessive sugar intake and to meet the 39 growing demand of consumers for a healthier lifestyle, the dairy industries have made use 40 of these new standards and concerns to manufacture products such as dulce de leche, 41 42 chocolate, ice cream and yogurt with low sugar content (Castanheira, 2012; McCain et 43 al., 2018; Moore et al., 2020). Nutritionally, dulce de leche (DL) has high energy value 44 and high concentration of proteins, minerals, fats and carbohydrates (BRASIL, 1997; 45 Francisquini et al., 2018; Stephani et al., 2019). Normally, the pans for the production of DL are simple to operate and maintain, being made of stainless steel with a double wall 46 47 through which the steam circulates and an agitator shaft. This has the purpose of preventing the portion of the liquid in direct contact with the equipment walls from 48 49 burning (modifying the sensory characteristic of the product) or forming incrustations (fouling) along the concentration in addition to causing the temperature to be homogenous 50 thus facilitating the evaporation of water. Upon reaching the desired point the product 51 52 must be cooled and packaged (Perrone et al., 2019; Stephani et al., 2019).

With the heat treatment of milk, two types of inlays can occur in the pan: type A (formed 53 54 mostly by proteins with a soft, white appearance) and type B (formed mostly from minerals, being more compact and grayer) (Bansal & Chen, 2006). As they differ in 55 composition, it is necessary to use different cleaning processes to efficiently remove each 56 type of deposit (Fickak et al., 2011; Goode et al., 2013; Jeurnink & Brinkman, 1994; 57 Morison & Thorpe, 2002). When adding sugar to milk, in the manufacture of DL, there 58 is a reduction in the occurrence of these incrustations, since there is an increase in the 59 60 surface tension of water and in the chemical potential of proteins, favoring the state of lower surface of the proteins (native protein). Moreover, the addition of sugar causes 61 62 rheological and kinetic changes that interfere with protein-protein interactions (Baier et 63 al., 2004; Kendrick et al., 1997). Thus, the traditional DL industry does not present considerably many cases of encrustation, but when it comes to dulce de leche with low 64 65 sugar content, changes in the product occur and they can lead to the formation of deposits 66 on the equipment. These fouling can result in loss of productivity, in loss of the thermal exchange efficiency, in pressure drop in the equipment and in increase of additional costs 67 related to cleaning products and energy (Gandhi et al., 2017; Tanguy et al., 2019; 2016). 68

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- 70 Material & methods
- 71

72 Deposit formation

Pasteurized whole milk used was purchased in the local market (Viçosa, Brazil). The temperature of the inlet product was 4 °C and added with different sucrose contents (0%, 5%, 10%, 15% and 20% (w·w⁻¹)). This process ensured the same protein content of all blends since for all the mixtures the same amount of milk was weighed and only then added the sucrose according to the desired percentages. The samples were concentrated in a laboratory scale atmospheric pressure evaporator (process simulator) (Vorwerk Thermomix TM5) nearly at 125 °C for 90 min with 1 g (centrifugal force) of rotation. The deposits formed were collected after the end of the concentration by scraping the surface of the equipment - mass of deposit - and weighed in analytical balance (BEL engineering® Mark 214). For the characterization of the deposits, was performed dehydration using the freeze-drying method (Labconco - Freezone 2.5 Plus) with temperature -88 °C and pressure 0.014 mbar. This method was used to standardize the samples.

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87 *Characterization of deposits*

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89 *Total protein*

90 Total nitrogen from each deposit formed from the different samples was determined by 91 using the micro Kjeldahl method, according the AOAC 982.38 method, it is an indirect 92 methodology because when considering a conversion factor of 6.38 it is possible to 93 estimate the protein contents (Wang *et al.*, 2016), the results were expressed in % dry 94 basis.

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96 *Moisture and dry matter*

97 The moisture content was measured by using 1.90 ± 0.05 g of each deposit and a 98 thermogravimetric balance (Sartorius® MA150). The total solids content was obtained 99 by percentage difference of the moisture content found and with this percentage is 100 possible to calculate the dry extract.

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102 *Carbon, Hydrogen and Nitrogen*

103 The Elemental Analysis was performed by means of an elemental analyzer (Perkin Elmer
104 2400 series ii) to determine the percentages of carbon, hydrogen, and nitrogen of the

samples of the deposits formed in the evaporator. This analysis is based on the PreglDumas method whose samples are subjected to combustion in an atmosphere of pure
oxygen and the gases resulting from this combustion are quantified in a TCD detector
(thermal conductivity detector).

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110 Calcium and Phosphorus

For the analysis of the trend of calcium and phosphorus content in the deposits it has been
used a semi-quantitative analysis by energy dispersive spectrophotometer – EDS
(HITACHI® TM3000).

And for quantitative analysis, the methods used were carried out according to the methodology of AOAC (2012), as official methods 985.35 using flame atomic absorption spectroscopy (FAAS) and 984.27 using inductively coupled plasma-atomic emission spectrometry (ICP-AES) for the calcium and phosphorus analyzes, respectively.

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119 Scanning electron microscopy

Samples of deposits were analyzed by scanning electron microscopy – SEM (HITACHI®
TM3000) with increase of 500x.

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123 FT-Raman spectroscopy

Fourier-transform Raman spectrometer (Bruker, RFS 100/S) with a germanium detector using liquid nitrogen as coolant and with 1064 nm excitation from a Nd:YAG laser was used to characterize the deposits according to their dispersive ability. A few milligrams of the sample were placed into a small aluminum sample cup, the laser light with a power of 20 mW was introduced and focused on the sample, then the scattered radiation was collected at 180°. For each spectrum an average of 512 scans were performed at a resolution of 4 cm⁻¹, over the 4000–50 cm⁻¹ range. The OPUS 6.0 (Bruker Optik,

| 131 | Ettlingen, | Germany) | software | program | was | used | for | Raman | data | acquisition | (Almeida, |
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132 Oliveira, Stephani & De Oliveira, 2011; Rodrigues Júnior et al., 2016).

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134 Results and discussion

135 *Concentration of Milk*

136 Corroborating with Figure 1, the same is seen after weighing the deposits, which goes 137 from 1.9 g to 21.5 g in the mix with 20% m·m⁻¹ of sucrose and without added sucrose, 138 respectively, which represents an increase of more than 11 times in incrustation.

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140 Physical-chemical characterization of deposits

When calculating the water-protein ratio of the deposits, it has been observed that with the decrease in the sucrose content there is a greater water retention by the proteins reaching approximately 4 g $H_2O\cdot g$ proteína⁻¹ corroborating with the literature (Walstra, Wouters, & Geurts, 2006).

145 It is still possible to say that the relationship between the lower content of deposits in the 146 production of traditional DL is not only related to the dilution of milk constituents by the 147 addition of sucrose, since the increase in deposits does not evolve in the same proportion 148 as the decrease of the dilution factor, thus indicating the protective effect of sucrose 149 amidst the concentration of the product (de Jong *et al.*, 1992).

Regarding the low content of calcium and phosphorus in the deposit resulting from the product with greater sucrose addition, it corroborates with the previous results and indicates that sucrose stabilizes milk proteins (Lee & Timasheff, 1981). As the relationship between calcium and phosphorus (Ca·P⁻¹) remains around 1.75, with no statistically significant difference by the Tukey test (p < 0.05) for different sucrose levels, this indicates that despite the increase in minerals in the deposit, these are precipitated in the same proportion, therefore, the phosphate ester groups of the caseins together with the bound calcium are assumed not to be part of the calcium phosphate matrix, thus beingof the tricalcium phosphate type (Gaucheron, 2005).

159 Pearson's correlation was used in order to observe whether the interactions between the analyzes performed and the variables of the experiment (added sucrose content, dilution 160 161 factors and concentration) were significant. Based on Table S1, it is possible to observe that the relationship between calcium and phosphorus does not present a statistically 162 163 significant correlation to the analyzed parameters. However, all other variables are 164 statistically significant in relation to the added sucrose content and to the dilution factor. But when analyzing in relation to the concentration factor, it is observed that the 165 166 percentage of total protein is not significant as it is in relation to the dilution factor, 167 therefore it is possible to conclude that the concentration, despite influencing proteins, the addition of sugar ends up generating a protective effect on them that is removed by 168 169 completely eliminating sucrose from milk.

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171 Scanning Electron Microscopy

The deposit observed in Figure A1 indicates Type A deposits (Burton, 1968), that is, with 172 173 high protein content, which goes with the other analyzes carried out and that show the 174 relationship between the decrease in sucrose and the increase in proteins (from 12.94 to 31.31% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposits with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposite with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis for deposite with the addition of 20 and 0% (w·w⁻¹) of protein on a dry basis fo 175 ¹) of sucrose, respectively) and also to the increase in moisture present in this type of 176 deposit (going from 18.43 to 54.23% humidity for deposits with addition of 20 and 0% 177 $(w \cdot w^{-1})$ sucrose, respectively), once the rough structure is able to retain more water than 178 179 the compact structure of the deposit formed in production of traditional DL. A change in the quantity and structure of the deposits with the addition of sucrose is also observed by 180 181 Zhang *et al.* (2018). Besides, this structure shows that the adopted cleaning procedures 182 must be different, since the protein deposit requires specific cleaning and specific chemical products in a certain sequence: first the alkaline solution for the removal of the
protein layer (top) and then the acidic solution for the removal of the innermost layer minerals (Jeurnink & Brinkman, 1994).

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187 *Raman spectroscopy*

In C2 (dulce de leche) and C3 (sucrose) spectra it is possible to perceive some of the main marking bands of sucrose, such as the band observed at 850 cm⁻¹ referring to the vibrational deformation mode of CH₂, the bands at 642 and 523 cm⁻¹, referring to the deformation modes of the glucofuran ring, and the 403 cm⁻¹ band, referring to a coupled mode to the sum of the deformation modes of the O39-C32-C31 bond and to the deformation of the C37-C31-O27 group of the glucopyran ring (Brizuela et al., 2012).

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Table legends:

Table 1:

290 Pearson correlations between the analyzed parameters and the sucrose content, dilution

291 factor and evaporation rate in the manufacture of DL.

| | Sucrose added (% w·w ⁻¹) | Dilution Factor | Evaporation Ratio |
|---|---|--------------------|----------------------|
| Deposit (% w·w ⁻¹) | -0.942 | -0.948 | 0.997 |
| Moisture (% w·w ⁻¹) | -0.953 | -0.952 | 0.793 |
| Total protein (% w·w ⁻¹) | -0.901 | -0.897 | 0.687 |
| Protein on dry basis (% w·w ⁻¹) | -0.957 | -0.956 | 0.815 |
| Nitrogen (%) | -0.953 | -0.945 | 0.846 |
| Calcium (norm. Wt. %) | -0.920 | -0.926 | 0.997 |
| Phosphorus (norm. Wt. %) | -0.902 | -0.906 | 0.987 |
| Ca·P ⁻¹ | -0.062 | -0.053 | -0.157 |
| Calcium (mg·100g ⁻¹) | -0.984 | -0.984 | 0.855 |
| Phosphorus (mg·100g ⁻¹) | -0.994 | -0.995 | 0.887 |
| Ca·P ⁻¹ | -0.691 | -0.683 | 0.386 |