Molecular identification of hymenopteran parasitoids and their endosymbionts from agromyzids — Supporting Information

Xuefen Xu, Ary A Hoffmann, Paul A Umina, Samantha E Ward, Marianne P Coquilleau, Mallik B Malipatil and Peter M Ridland (2023)

Supplementary Figures S1-S10

Scanning electron micrograph (SEM) and photomicrographs for:

	Page No
Fig. S1 Aprostocetus sp.	2
Fig. S2 Asecodes sp.	4
Fig. S3 Closterocerus mirabilis	8
Fig. S4 Chrysocharis pubicornis	10
Fig. S5 Neochrysocharis okazakii	13
Fig. S6 Trigonogastrella parasitica	17
Fig. S7 Dacnusa areolaris	20
Fig. S8 Opius cinerariae	23
Fig. S9 Opius sp. 1	26
Fig. S10 Opius sp. 2	29
References	34

SEM and photomicrographs for *Neochrysocharis formosa* are given in Xu *et al.* (2022).

Terminology of characters for chalcidoids follows Gibson (1997), Gibson *et al.* (1998) and Hansson (2022); and for braconids follows van Achterberg (1993), Sharkey and Wharton (1997), Wharton (2006) and Karlsson and Ronquist (2012).

1. Aprostocetus sp. Westwood, 1833

(Hymenoptera: Eulophidae: Tetrastichinae)

Diagnostic characters (Bouček 1988; Reina & La Salle, 2006)

i. Fronto-facial sutures near to anterior ocellus (Fig. S1-A)

ii. Malar sulcus present and straight (Fig. S1-B)

iii. Cercal setae unequal in length, with one being distinctly longer than the others and sinuate (Fig. S1-C)

iv. Mesoscutellum with 2 pairs of setae and with 2 pairs of longitudinal lines (Fig. S1-D).

v. Propodeum with median carina (MC) present with a raised lobe of the callus, which partially overhangs the outer rim of the spiracles (Fig. S1-D).

vi. Mid-lobe of mesoscutum with median line and a single row of adnotaular setae just mesal to the notaulus. (Fig. S1-E).

vii. Flagellum with 3 funiculars and 3 clavomeres (club) (Fig. S1-G); type 2 peg sensilla (Fig. S1-F).

viii. Postmarginal vein (PMV) absent or less than 0.5 the length of stigmal vein (STV) (Fig. S1-H).

ix. Three or more setae on submarginal vein (SMV) (Fig. S1-H).





Figure S1. Aprostocetus sp. (A) \bigcirc head; (B) \bigcirc malar sulcus; (C) \bigcirc longest cercal setae; (D) \bigcirc scutellum; (E) \bigcirc mesoscutum; (F) \bigcirc peg sensilla on antenna; (G) \bigcirc antenna; (H) \bigcirc forewing. All scale bars are 50 µm except (F) is 4 µm. AS=adnotaular setae; MC=median carina; MS=malar sulcus; PMV=postmarginal vein; SMV=submarginal vein; STV=stigmal vein.

2. Asecodes sp. Förster, 1856

(Hymenoptera: Eulophidae: Entedoninae: Entedonini)

To date, all *Asecodes* spp. reared from agromyzids were originally considered to be in the genus *Teleopterus* (Hansson 1994). Bouček (1988) noted 3-4 species of *Teleopterus* in Australian collections, but *T. atripes* was the only one species described (Girault, 1915a; Dahms 1983). Subsequently, *Teleopterus* was synonymised with *Asecodes* (Hansson 1996). Gumovsky (2001) then synonymised *Asecodes*, *Neochrysocharis*, *Hispinocharis* and *Mangocharis* with *Closterocerus*. However, molecular analysis led Burks *et al.* (2011) to remove *Neochrysocharis* and *Asecodes* from synonymy.

Belongs to Entedonini - two setae dorsally on SMV (Fig. S2-I), one pair of setae on mesoscutellum (Fig. S2-C).

Diagnostic characters (Bouček 1988; Hansson 1994, 1996; Fisher et al. 2005; Burks 2019)

i. Subtorular grooves present (SG); fronto-facial suture distinctly separated from anterior ocellus and V or Y-shaped (Fig. S2-A).

ii. Head with distinct occipital median furrow (OF) (Fig. S2-B).

iii. Mesosoma slightly sculptured; pronotum very short and without transverse carina; notauli are incomplete; mesoscutum and scutellum without longitudinal grooves; propodeum smooth, shiny and without median carina and plicae. (Fig. S2-C; Fig. S2-D)

iv. Petiole not distinct (Fig. S2-E).

v. Flagellum with 2 funiculars and 3 clavomeres (Fig. S2-G, Fig. S2-H); σ flagellum with setae confined to a basal whorl on each segment (Fig. S2-H); (but Hansson (1994) noted that in some genera of Entedoninae, this character is present only in some species, so it is not a suitable character to define limits for a genus); type 2 peg sensilla on flagellum (Fig. S2-F).

vi. Forewing with 2 indistinct stigmal hair lines (rows of setae radiating from STV; PMV shorter than STV (Fig. S2-I).





Figure S2. Asecodes sp. (A) \bigcirc head; (B) \bigcirc occipital grooves; (C) \bigcirc mesosoma, dorsal view; (D) \bigcirc mesosoma, lateral view; (E) \Diamond petiole; (F) \bigcirc type 2 peg sensilla on

antenna; (G) \bigcirc antenna; (H) \bigcirc antenna; (I) \bigcirc forewing. All scale bars are 50 µm except (F) is 4 µm. OF=occipital median furrow; PMV=postmarginal vein; SG=subtorular grooves; SMV=submarginal vein; STV=stigmal vein.

3. Closterocerus mirabilis Edwards & La Salle, 2004

(Hymenoptera: Eulophidae: Entedoninae: Entedonini)

Belongs to Entedonini - two setae dorsally on SMV (Fig. S2-I), one pair of setae on mesoscutellum (Fig. S2-C).

Diagnostic characters (Edwards & La Salle, 2004)

i. Frontal-facial suture V-shaped, connecting to scrobal grooves but not connected to each other medially; short subtorular grooves present (Fig. S3-A).

ii. Mesosoma: Pronotum without a distinct transverse carina. The notauli on the mesoscutum are incomplete (Fig. S3-C, Fig. S3-D); pronotum, mesoscutum and mesoscutellum strongly and evenly reticulate dorsally and laterally, except posterior margin of mesoscutellum with transverse smooth band (Fig. S3-B, Fig. S3-C, Fig. S3-D, Fig. S3-E); mesoscutellum with a single pair of setae (Fig. S3-E).

iii. Petiole not distinct (Fig. S3-F).

iv. Antenna compressed, scape widest at apex; flagellum with 2 funicular segments (wider than long) and club 3-segmented. (Fig. S3-G).

v. Forewing with a characteristic infuscate pattern: a transverse stripe at the wing apex, a complete dark transverse stripe at level of stigmal vein, a cloud beneath the marginal vein, and a stripe along the hind margin of the wing. Speculum is large. Cubital row of setae complete to basal vein. Single hairline from stigmal vein. Radial cell bare. (Fig. S3-H).





Figure S3. Closterocerus mirabilis. (A) \bigcirc head; (B) \bigcirc mesosoma, lateral view; (C) \bigcirc mesosoma, dorsal view.; (D) \bigcirc mesoscutum; (E) \bigcirc mesoscutellum and propodeum (F) \bigcirc metasoma, showing petiole. (G) \bigcirc antenna; (H) \heartsuit forewing; All scale bars are 50 µm. PMV=postmarginal vein; SMV=submarginal vein; STV=stigmal vein.

4. Chrysocharis pubicornis (Zetterstedt, 1838)

(Hymenoptera: Eulophidae: Entedoninae: Entedonini)

Belongs to Entedonini - one pair of setae on mesoscutellum (Fig. S4-C); two setae dorsally on SMV (Fig. S4-F),

Diagnostic characters (Hansson 1985; Ikeda 1996; Narendran et al. 2013)

i. Head without a sulcus surrounding ocellar triangle; fronto-facial suture distinctly separated from anterior ocellus and usually V or Y-shaped (Fig. S4-A).

ii. Type 1 peg sensilla on Q antennae. Third annellus enlarged (Fig. S4-B, Fig. S4-G)

iii. Mesosoma slightly to strongly sculptured; pronotum with a low carina; notauli are incomplete; mesoscutum and mesoscutellum without longitudinal grooves; mesoscutellum with a single pair of setae (Fig. S4-C; Fig. S4-D).

iv. Propodeum with raised structure on anterior part (Fig. S4-E); petiole shorter than propodeum, wider than long (Fig. S4-E).

v. Relative measurements for length of marginal/ postmarginal/ stigmal veins (MV/PMV/STV): 7.1/2.1/1.0 (Hansson 1985) (Fig. S4-F).





Figure S4. Chrysocharis pubicornis. (A) \bigcirc head; (B) \bigcirc peg sensillae on antenna; (C) \bigcirc ; mesosoma, dorsal view; (D) \bigcirc mesosoma, lateral view; (E) \bigcirc propodeum; (F) \bigcirc forewing; (G) \bigcirc antenna. All scale bars are 50 µm. MV=marginal vein; PMV=postmarginal vein; SMV=submarginal vein; STV=stigmal vein.

5. Neochrysocharis okazakii Kamijo, 1978

(Hymenoptera: Eulophidae: Entedoninae: Entedonini)

Belongs to Entedonini - two setae dorsally on SMV (Fig. S5-I), one pair of setae on mesoscutellum (Fig. S5-B, S5-C)

Diagnostic characters (Kamijo 1978; Hansson 1990; Fisher & La Salle 2005; Fisher et al. 2005)

i. Head with fronto-facial suture distinctly separated from anterior ocellus and V or Y-shaped (Fig. S5-A).

ii. Pronotum without transverse carina (Fig. S5-B); mesoscutum and scutellum without longitudinal grooves and notauli are incomplete (Fig. S5-D); transepimeral sulcus (tps) curved postero-dorsally (Fig. S5-E).

iii. propodeum smooth, shiny and without median carina and plica (Fig. S5-F); callus with 2 setae; petiole not distinct (Fig. S5-F); mesosoma and gaster relatively smooth and shiny, without distinct reticulate sculpture. (Fig. S5-D, S5-H).

iv. Flagellum with 2 funiculars and 3 fused clavomeres (Fig. S5-G, S5-H).

v. PMV present and shorter than STV. No infumation associated with stigmal vein. Forewing about 2.0 times longer than wide, using the HW/LW definition given by Hansson (1990) (LW=length of forewing, measured from base of marginal vein to the place on the wing farthest away from base of marginal vein; HW=height of forewing, represented by the line right-angular to the line representing LW) (Fig. S5-I).





Figure S5. Neochrysocharis okazakii. (A) $\[mathcar{P}\]$ head; (B) $\[mathcar{P}\]$ mesosoma, dorsal view; (C) $\[mathcar{O}\]$ mesoscutum; (D) scutellum and propodeum; (E) $\[mathcar{P}\]$ mesosoma, lateral view; (F) $\[mathcar{P}\]$ gaster. (G) $\[mathcar{O}\]$ antenna; (H) $\[mathcar{P}\]$ antenna; (I) $\[mathcar{O}\]$ forewing. All scale bars are 50 µm. SMV=submarginal vein; PMV=postmarginal vein; STV=stigmal vein.

6. Trigonogastrella parasitica Girault, 1915

(Hymenoptera: Pteromalidae: Pteromalinae)

Diagnostic characters (Girault 1915b; Bouček 1988; Fisher et al. 2005)

i. Forewing with PMV and STV well-developed; PMV elongate, as long as the MV; the STV distinctly shorter than both MV and PMV, PMV twice as long as STV (Fig. S6-G).

ii. Flagellum with 6 funiculars, 3 clavomeres; two annelli (Fig. S6-H).

iii. Mandibles four-dentate (Fig. S6-A).

iv. No post-occipital carina (Fig. S6-B).

v. Mesosoma and propodeum punctate; propodeum with a complete median carina and straight spiracular sulci (Fig. S6-C; Fig. S6-D).

vi. Petiole present and distinct (Fig. S6-E).

vii. Second gastral tergite shorter than first tergite; first gastral tergite not broadly concave (Fig. S6-I).

viii. Scape of σ is entirely light brown, with the flagellum dark brown (Fig. 6-F). In contrast, the only the basal part of the scape of φ is light brown. The clava of the φ is more distinct than the clava of the σ (Fisher *et al.* 2005).

https://keyserver.lucidcentral.org/key-server/data/0b0d070f-0c0f-490f-8201-0f04050f0908/media/Html/trigonogastrella.html





Figure S6. Trigonogastrella parasitica. (A) $\stackrel{\circ}{\rightarrow}$ head; (B) $\stackrel{\circ}{\rightarrow}$ occiput; (C) $\stackrel{\circ}{\rightarrow}$ mesosoma, dorsal view; (D) $\stackrel{\circ}{\rightarrow}$ mesosoma, lateral view; (E) $\stackrel{\circ}{\rightarrow}$ petiole; (F) $\stackrel{\circ}{\rightarrow}$ scape; (G) $\stackrel{\circ}{\rightarrow}$ forewing; (H) $\stackrel{\circ}{\rightarrow}$ antenna; (I) $\stackrel{\circ}{\rightarrow}$ gaster. All scale bars are 50 µm. PMV=postmarginal vein; STV=stigmal vein.

7. Dacnusa areolaris (Nees ab Esenbeck, 1812)

(Hymenoptera: Braconidae: Alysiinae)

Diagnostic characters (Griffiths 1966; Wharton & Austin 1991; Fisher *et al.* 2005; Berry 2007)

1. Mandibles exodont - pointing outwards and not coming close to meeting medially (Fig.S7-A).

2. Mandible with 3 teeth; tooth 1 and 3 rounded, tooth 2 long and acutely pointed (Fig.S7-B).

3. Mesonotum, metapleuron, and petiole densely setose (Fig. S7-C; S7-D)

4. Forewing: stigma long and narrow, at least 10 times longer than wide; vein r arising in basal 1/5 of stigma (Fig. S7-E).

5. Hindwing: 1st sub-basal cell slightly more than 1/2 the length of the basal cell (Fig. S7-F).





Figure S7. Dacnusa areolaris. (A) $\stackrel{\circ}{\rightarrow}$ head; (B) $\stackrel{\circ}{\rightarrow}$ mandible; (C) $\stackrel{\circ}{\rightarrow}$ mesonotum; (D) $\stackrel{\circ}{\rightarrow}$ lateral view of body showing metapleural setae; (E) $\stackrel{\circ}{\rightarrow}$ forewing; (F) $\stackrel{\circ}{\rightarrow}$ hindwing. All scale bars are 50 µm.

8. Opius cinerariae Fischer 1963

(Hymenoptera: Braconidae: Opiinae)

Diagnostic characters (Belokobylskij et al. 2004)

i. Gap between clypeus and mandible absent or narrow (Fig. S8-A, Fig. S8-B) when mandibles are closed.

ii. The malar suture is absent (Fig. S8-C).

iii. Precoxal sulcus (PCS) deep and distinctly sculptured (Fig. S8-D).

iv. Mesonotum with distinct median pit (MP) and the notauli (NO) are sculptured over the anterior half of the mesoscutal disc (Fig. S8-E).

v. Propodeum rugose (Fig. S8-F).

vi. First metasomal tergite densely striate; second metasomal tergite very finely coriaceous; remaining tergites smooth (Fig. S8-G, Fig. S8-H, Fig. S8-I, Fig. S8-J)

vii. The radial cell of forewing is shortened (Fig. S8-K)

viii. Nervellus vein (cu-a) and submedial (sub-basal) cell of hind wing absent (Fig. S8-L).





Figure S8. Opius cinerariae. (A) \Im head, frontal view; (B) \Im clypeus and mandibles showing hypoclypeal cavity; (C) \Im head, lateral view (no malar sulcus); (D) \Im mesopleuron; (E) \Im mesonotum showing median pit and notauli; (F) \Im rugose propodeum; (G) \Im metasoma (dorsal); (H) \Im metasoma (ventral); (I) \Im metasoma

(dorsal); (J) \circlearrowright metasoma (ventral); (K) \updownarrow forewing; (L) \circlearrowright hindwing. All scale bars are 50 µm. HC =hypostomal cavity; NO =notaulus; MP =median pit on mesonotum; PCS=precoxal sulcus.

9. Opius sp. 1

(Hymenoptera: Braconidae: Opiinae)

Opius sp.1 is identical to *Opius* morphospecies 1 (Coquilleau 2020). It is very close to *Opius atricornis* Fischer (Belokobylskij *et al.* 2004).

i. Clypeus narrow and labrum broadly exposed below clypeus (Fig. S9-A, Fig. S9-B) when mandibles are closed.

ii. The malar suture is absent (Fig. S9-C).

iii. Precoxal sulcus indistinct (Fig. S9-D).

iv. Mesonotum without distinct median pit and the notauli are not sculptured over the anterior half of the mesoscutal disc (Fig. S9-E).

v. Propodeum unsculptured and not rugose (Fig. S9-F).

vi. First metasomal tergite is not densely striate; second and third metasomal tergites are smooth and polished (Fig. S9-G, Fig. S9-H, Fig. S9-I, Fig. S9-J)

vi. The radial cell of the forewing is not shortened cf. *O. cinerariae*. The 3M vein is tubular and extends nearly to the margin of the wing; the 2CUb vein is visible for half of the distance to the margin of the wing (Fig. S9-K).

vii. Nervellus vein (cu-a) and submedial (sub-basal) cell of hind wing absent (Fig. S9-L).

viii. Body and head burnt orange, majority of the mesoscutum shiny black, with some variation to a lighter brown colour. Antennae brown, proximal half of the scape burnt orange. Coxae and legs, first and second metasomal tergites are pale orange, yellowish. Remaining tergites are brown. (Fig. S9-M)







Figure S9. Opius sp. 1. (A) \Im head, frontal view; (B) \Im clypeus narrow and crescent shaped, mandibles, hypoclypeal cavity; (C) \Im head, lateral view; (D) \Im mesopleuron; I \Im mesonotum; (F) \Im propodeum unsculptured; (G) \Im metasoma (dorsal); (H) \Im metasoma (ventral); (I) \Im metasoma (dorsal); (J) \Im metasoma (ventral); (K) forewing; (L) hindwing; (M) \Im habitus. All scale bars are 50 µm.

10. Opius sp. 2

(Hymenoptera: Braconidae: Opiinae)

Opius sp. 2 is identical to *Opius* morphospecies 2 (Coquilleau 2020). It differs from *Opius* sp. 1 primarily by colour and the transparency of some veins in the forewing.

i. Clypeus narrow and labrum broadly exposed below clypeus (Fig. S10-A, Fig. S10-B) when mandibles are closed.

ii. The malar suture is absent (Fig. S10-C).

iii. Precoxal sulcus indistinct (Fig. S10-D).

iv. Mesonotum without distinct median pit and the notauli are not sculptured over the anterior half of the mesoscutal disc (Fig. S10-E).

v. Propodeum unsculptured and not rugose (Fig. S10-F).

vi. second and third metasomal tergites are smooth and polished (Fig. S10-G, Fig. S10-H, Fig S10-I, Fig. S10-J).

vi. The radial cell of forewing is not shortened cf. *O. cinerariae*. The 3M vein is nebulous and foreshortened; the 2CUb vein is nebulous (Fig. S10-K).

vii. Nervellus vein (cu-a) and submedial (sub-basal) cell of hind wing absent (Fig. S10-L).

viii. Body shiny black. Antennae and scape brown. Coxae and legs yellowish. First mesosomal tergite with light brown tinge, remaining tergites brown nearly black (Fig. S10-M)







Figure S10. Opius sp. 2. (A) $\[mathbb{P}\]$ head, frontal view; (B) $\[mathbb{P}\]$ clypeus narrow and crescent shaped, mandibles, hypoclypeal cavity; (C) $\[mathbb{O}\]$ head, lateral view (no malar sulcus); (D) $\[mathbb{O}\]$ mesopleuron; (E) $\[mathbb{O}\]$ mesonotum (no medial pit); (F) $\[mathbb{O}\]$ propodeum; (G) $\[mathbb{P}\]$ metasoma (dorsal); (H) $\[mathbb{P}\]$ metasoma (ventral); (I) $\[mathbb{O}\]$ metasoma (dorsal); (J) $\[mathbb{O}\]$ metasoma (ventral); (K) $\[mathbb{P}\]$ forewing; (L) $\[mathbb{P}\]$ hindwing; (M) $\[mathbb{O}\]$ habitus. All scale bars are 50 µm.

References

- Belokobylskij SA, Wharton RA and La Salle J (2004) Australian species of the genus Opius Wesmael (Hymenoptera: Braconidae) attacking leaf-mining Agromyzidae, with the description of a new species from South-east Asia. Australian Journal of Entomology 43, 138–147.
- **Berry JA** (2007) Alysiinae (Insecta: Hymenoptera: Braconidae). Fauna of New Zealand.
- **Bouček Z** (1988) Australasian Chalcidoidea (Hymenoptera). A biosystematic revision of genera of fourteen families, with a reclassification of species. CAB International, Wallingford.
- Burks RA (2019) Key to the Nearctic genera of Eulophidae, subfamilies: Entedoninae, Euderinae, and Eulophinae (Hymenoptera: Chalcidoidea). Available from: <u>https://faculty.ucr.edu/~heraty/Eulophidae/index.html</u> Accessed 20 October 2022].
- Burks RA, Heraty JM, Gebiola M and Hansson C (2011) Combined molecular and morphological phylogeny of Eulophidae (Hymenoptera: Chalcidoidea), with focus on the subfamily Entedoninae. *Cladistics* 27, 581-605.
- **Coquilleau MP** (2020) Seasonality and community composition of parasitoid wasps of four agromyzid leafminer species (Diptera: Agromyzidae) in Victoria. MPhil Thesis, The University of Melbourne. Available from URL: <u>https://minerva-</u> access.unimelb.edu.au/handle/11343/252730 [Accessed 20 October 2022]
- Dahms EC (1983) A checklist of the types of Australian Hymenoptera described by Alexandre Arsene Girault: II. Preamble and Chalcidoidea species A-E with advisory notes. *Memoirs of the Queensland Museum* 21, 1–255.
- Edwards CM and La Salle J (2004) A new species of *Closterocerus* Westwood (Hymenoptera: Eulophidae), a parasitoid of serpentine leafminers (Diptera: Agromyzidae) from Australia. *Australian Journal of Entomology* 43, 129–132.
- Fischer M (1963) Neue Zuchtergebnisse von Braconiden (Hymenoptera). Zeitschrift für Angewandte Zoologie 50, 195–214.

- Fisher N and La Salle J (2005) A new species of *Neochrysocharis* Kurdjumov (Hymenoptera: Eulophidae), a parasitoid of serpentine leafminers (Diptera: Agromyzidae) in Southeast Asia. *Zootaxa* 1044, 27–34.
- Fisher N, Ubaidillah R, Reina P and La Salle J (2005) *Liriomyza* parasitoids in Southeast Asia. Available from: <u>https://keys.lucidcentral.org/keys/v3/Liriomyza/index.html [Accessed 20 October 2022].</u>
- Gibson GAP (1997) Chapter 2. Morphology and Terminology, In Gibson GAP, Huber JT and Woolley JB (eds), Annotated keys to the genera of Nearctic Chalcidoidea (Hymenoptera). National Research Council of Canada, NRC Research Press, Ottawa, Canada, pp 16–44.
- Gibson GAP, Read JD and Fairchild R (1998) Chalcid wasps (Chalcidoidea): illustrated glossary of positional and morphological terms. Available from: <u>http://www.canacoll.org/Hym/Staff/Gibson/apss/chglintr.htm</u> [Accessed 20 August 2022].
- Girault AA (1915a) Australian Hymenoptera Chalcidoidea IV. Supplement. Memoirs of the Queensland Museum 3, 180–299.
- **Girault AA** (1915b) Australian Hymenoptera Chalcidoidea VI. Supplement. *Memoirs of the Queensland Museum* **3**, 313–346.
- Griffiths GCD (1966) The Alysiinae (Hym. Braconidae) parasites of the Agromyzidae (Diptera). III. The parasites of *Paraphytomyza* Enderlein, *Phytagromyza* Hendel and *Phytomyza* Fallén. *Beiträge zur Entomologie* 16, 775–951
- Gumovsky A (2001) The status of some genera allied to *Chrysonotomyia* and *Closterocerus* (Hymenoptera: Eulophidae, Entedoninae), with description of a new species from Dominican amber. *Phegea* 29, 125–141.
- Hansson C (1985) Taxonomy and biology of the Palearctic species of *Chrysocharis*Förster, 1856 (Hymenoptera: Eulophidae). *Entomologica Scandinavica Supplement* 26, 1–130.

- Hansson C (1990) A taxonomic study on the Palearctic species of *Chrysonotomyia* Ashmead and *Neochrysocharis* Kurdjumov (Hymenoptera: Eulophidae). *Entomologica Scandinavica* 21, 29-52.
- Hansson C (1994). The classification of *Chrysonotomyia* Ashmead and *Telopterus* Silvestri (Hymenoptera: Eulophidae), with a review of the species in the Nearctic region. *Proceedings of the Entomological Society of Washington* 96, 665–673.
- Hansson C (1996) The status of the genera Asecodes Förster, Ionympha Graham and Teleopterus Silvestri (Hymenoptera: Eulophidae), with a review of Nearctic species. Entomologica Scandinavica 27, 159–167.
- Hansson C (2022) Terminology in Neotropical Eulophidae. Available from: <u>http://www.neotropicaleulophidae.com/Terminology1.html</u> [Accessed 20 October 2022].
- Ikeda E (1996) Revision of the Japanese Species of Chrysocharis (Hymenoptera, Eulophidae), III. Japanese Journal of Entomology 64, 551–569
- Kamijo K (1978) Chalcidoid parasites (Hymenoptera) of Agromyzidae in Japan, with description of a new species. *Kontyû, Tokyo* **46**, 455–469.
- Karlsson D and Ronquist F (2012) Skeletal morphology of *Opius dissitus* and *Biosteres carbonarius* (Hymenoptera: Braconidae), with a discussion of terminology. *PLOS ONE* 7(4), e32573. https://doi.org/10.1371/journal.pone.0032573
- Narendran TC, Razak N and Sureshan PM (2013) A taxonomic review of Chrysocharis Foerster (Hymenoptera: Eulophidae) of Indian subcontinent. Records of the Zoological Survey of India 113, 13-34.
- Reina P and La Salle J (2003) Key to the world genera of Eulophidae parasitoids (Hymenoptera) of leafmining Agromyzidae (Diptera). Available from URL: https://keys.lucidcentral.org/keys/v3/eulophidae_parasitoids/index.html [Accessed 20 October 2022].
- Sharkey MJ and Wharton RA (1997) Morphology and Taxonomy. Pages 19–38. In Wharton RA, Marsh PM and Sharkey MJ (eds) *Manual of the New World genera*

of Braconidae (Hymenoptera). Special Publication of the International Society of Hymenopterists **1**: 1–439.

van Achterberg C (1993) Illustrated key to the subfamilies of the Braconidae (Hymenoptera: Ichneumonoidea). Zoologische Verhandelingen Leiden 283, 1– 189. See also WWW version Available from: <u>https://braconidae.linnaeus.naturalis.nl/linnaeus_ng/app/views/introduction/topic.</u> <u>php?id=2935</u> [Accessed 20 October 2022].

Wharton RA (2006) The species of *Sternaulopius* Fischer (Hymenoptera: Braconidae, Opiinae) and the braconid sternaulus. *Journal of Hymenoptera Research* 15, 317–347.

Wharton RA and Austin AD (1991) Revision of Australian Dacnusini (Hymenoptera: Braconidae: Alysiinae), parasitoids of cyclorrhaphous Diptera. *Australian Journal of Entomology* 30, 193–206.

Xu X, Hoffmann AA, Umina PA, Coquilleau MP, Gill A and Ridland PM (2022)

Identification of two leafminer parasitoids (Hymenoptera: Eulophidae),

Neochrysocharis formosa and *Proacrias* sp. from Australia, with both showing thelytoky and infection by *Rickettsia*. *Austral Entomology* **61**, 358–369.