The death of Kaakutja: a case of peri-mortem weapon trauma in an Aboriginal man from north-western New South Wales, Australia

Michael Westaway^{1,*}, Douglas Williams², Richard Wright³, Rachel Wood⁴, Jon Olley⁵, Jaime Swift⁶, Sarah Martin⁷, Justine Kemp⁵, Shane Rolton⁸ & William Bates⁹

 ¹Environmental Futures Research Institute, Griffith University, 170 Kessels Road, Nathan, QLD 4111, Australia
²Access Archaeology & Heritage, PO Box 816, Moruya, NSW 2537, Australia
³Faculty of Arts and Social Sciences, University of Sydney, Sydney, NSW 2006, Australia
⁴Research School of Earth Sciences, The Australian National University, 142 Mills Road, Acton, ACT 2601, Australia
⁵Australian Rivers Institute, Griffith University, 170 Kessels Road, Nathan, QLD
4111, Australia
⁶College of Asia and the Pacific, The Australian National University, Canberra, ACT 0200, Australia
⁷Office of Environment & Heritage, 183 Argent Street, Broken Hill, NSW 2880, Australia
⁸Wysiwyg 3D, Unit 1, 22 Norman Street, Peakhurst, NSW 2210, Australia
⁹Paakantji Aboriginal Cultural Group, Broken Hill, NSW, Australia

* Author for correspondence (Email: m.westaway@griffith.edu.au)

Skeletal remains from a burial in New South Wales exhibit evidence of fatal trauma, of a kind normally indicative of sharp metal weapons, yet the burial dates to the mid thirteenth century—600 years before European settlers reached the area. Could sharp-edged wooden weapons from traditional Aboriginal culture inflict injuries similar to those resulting from later, metal blades? Analysis indicates that the wooden weapons known as 'Lil-lils' and the fighting boomerangs ('Wonna') both have blades that could fit within the dimensions of the major trauma and are capable of having caused the fatal wounds.

Keywords: Australia, Aboriginal, skeletal remains, burial, trauma, boomerang

Radiocarbon dating

Two samples taken from the skeletal remains were extracted for radiocarbon dating, a metatarsal from the left foot and one incisor. In addition, a yabby gastrolith extracted from the preserved stomach contents and a leaf compressed against the skull, and thought to possibly represent foliage incorporated in the original burial ceremony, were also dated.

Following physical cleaning and pre-treatment, samples were converted to carbon dioxide by combustion in a sealed tube (collagen and leaf) or reaction with phosphoric acid (gastrolith) before graphitisation over an iron catalyst in the presence of hydrogen and measurement in a single stage AMS (Fallon *et al.* 2010). All dates in this paper have been calibrated against SHCal13 (Hogg *et al.* 2013) or Bomb 13 SH 1_2 (Hua *et al.* 2013) in OxCal v.4.2 (Bronk Ramsey 2009). The metatarsal was dated three times. The first collagen extract was run twice (SANU-40414 and SANU-40505) and then a second collagen extraction was done to check for contamination (SANU-40822). Collagen extracted from the tooth was dated once. All four results are statistically identical (χ^2 -test: df=3, T=0.9 (5% 7.8)), yielding a date of AD 1260–1280 (95.4% confidence). The yabby gastrolith yielded a date a little later than the bone, of AD 1440–1615 (95.4% confidence) and the leaf provided a post-bomb radiocarbon signature; the calibrated date being AD 1956–1957 (see Table S1 below for all results).

The radiocarbon date on the leaf is regarded as problematic. The sample was extremely delicate, making separation of plant material and sediment difficult. The %C of the final sample is only 22%, substantially lower than normally expected of plant material (around 40%) suggesting the presence of non-plant material. Likewise, we have no quality assurance indicators to identify whether the carbonate within the gastrolith has recrystallised, and so it is not possible to confirm whether the date may be affected by contamination. In contrast, collagen from the bone and tooth dentine was well preserved and meets expected quality assurance criteria (van Klinken 1999). It is possible that this individual ate a substantial amount of protein from freshwater resources, which could feasibly contain a radiocarbon reservoir, as groundwater is known to enter river systems around Bourke (Meredith *et al.* 2009), making this date older than the age of the individual. Unusual δ^{13} C and elevated δ^{15} N values can sometimes be used to identify the consumption of freshwater protein (e.g. Cook *et al.*

2

2001; Wood *et al.* 2013). In northern NSW, a combination of aridity and C₄ grasses means both δ^{13} C and δ^{15} N are unusually elevated in the terrestrial food chain (average of three modern *Macropus* from Bourke, δ^{13} C -16.3±1.1‰, δ^{15} N 7.6±1.3‰; Fraser 2007), and so without further study of the freshwater stable isotope ecology it is not possible to ascertain whether a large proportion of the carbon within the collagen extracted is derived from carbon from freshwater resources.

Table S1. ¹⁴C dates taken directly from Kaakutja's skeletal remains, a yabby gastrolith from his stomach and a leaf from the sediment within the burial, thought initially to be possibly associated with the burial. Dates are calibrated against SHCal13 (Hogg *et al.* 2013) or Bomb 13 SH 1_2 (Hua *et al.* 2013) in OxCal v.4.2 (Bronk Ramsey 2009). Bone should have >1% collagen yield, >30% C and a C:N ratio of 2.9–3.4 (van Klinken 1999).

Sample information			AMS results			Quality assurance data and IRMS results						
		Pre-										
		treatment										
		(see			Calibrated date	Pretreat	Pretreat					
	Sample	comments	δ ¹³ C		(95.4%	yield	yield					
S-ANU	name	below)	(±1)	¹⁴ C age	confidence)	(mg)	(%)	$\delta^{13}C$	$\delta^{15}N$	%C	C:N	Comments
40414	foot	1	-10	745±20 BP	AD 1260–1280	18.8	11.5	-14.4	11.7	44.2	3.2	First collagen
40505	(sample 1)		-26	740±20 BP								extract
40822	foot	1	-13	750±22 BP		14.3	8.8	-14.0	11.3	45.6	3.3	Second
	(sample 2)											collagen
												extract
40821	incisor	1	-16	765±19 BP	AD 1220–1280	2.7	3.3	-15.0	11.8	44.3	3.3	
40820	gastrolith	2	-8	405±20 BP	AD 1440–1615	102.3	86.9			7^4		
41306	leaf	3	-29	102.10±0.2	AD 1956–1957	2.4	14.7			22^{4}		
				3 pmC								

Comments:

After physical cleaning of the sample, ultrafiltration pre-treatment consisted of washing in chloroform:methanol (2:1 ratio, room temperature (RT), 1 hr, subsequently dried), HCl (0.5M, 5°C, overnight), NaOH (0.1M, RT, 30 min), HCl (0.5M, RT, 1 hour), and then gelatinisation HCl (0.001M, 70°C, 20 hrs), EezeTM filtration and Ultrafiltration (precleaned VivaspinTM Turbo15 30 kDa MWCO). A second aliquot of extracted collagen was used for analysis within a Sercon 20-22 isotope ratio mass spectrometer (IRMS) connected to an ANCA elemental analyser (EA) operating in continuous flow mode, using an in-house gelatin reference and USGS-40 and USGS-41 international standards.

- 2. After physical cleaning of the surface of the sample, the gastrolith was leached in 0.1M HCl at 80 °C until at least 10% of the sample weight was lost.
- 3. After gentle physical cleaning of the surface of the sample, the leaf was washed in HCl (1M, 70 °C, 30 min), NaOH (1M, 70 °C, 1 hour, replaced until solution colourless), HCl (1M, 70 °C, 30 min). The sample was judged too delicate to bleach.
- 4. %C calculated volumetrically during gas collection for graphitisation. Both values are low. The gastrolith probably contained significant amounts of organic carbon and the leaf was extremely fragile, making sediment difficult to physically remove prior to analysis.

Optical dating

The samples were processed to isolate pure extracts of $180-212\mu m$ light-safe quartz grains. Sample processing followed standard procedures (e.g. Aitken 1998) and single-grain equivalent dose (D_e) values were determined using the modified single aliquot-regenerative dose (SAR) protocol of Olley *et al.* (2004), in combination with the acceptance/rejection criteria provided in Pietsch (2009).

The age modelling approach and estimates of dose rates followed standard procedures (Mejdahl 1979; Murray *et al.* 1987; Galbraith & Laslett 1993; Prescott & Hutton 1994; Galbraith *et al.* 1999; Roberts *et al.* 2000; Stokes *et al.* 2003) and produced a dose rate for the pit side sample estimated at 1.75 ± 0.13 Gy/ka. The single-grain D_e estimates for two samples are displayed in radial plots in Figure S1. Both samples are over-dispersed and in each case more than one dose population is evident. This is indicative of partial or heterogeneous bleaching; consequently, we have used the lowest dose population of grains to determine both the deposition age and the timing of the burial (see Olley *et al.* 2004). The lowest dose population in the sample collected from the side of the excavation pit has a D_e of 1.86 ± 0.10 Gy, which gives a deposition date of between AD 835 and 1055. The lowest dose population in the sample collected from inside the skull has a D_e of 1.05 ± 0.18 Gy applying the same dose rate, giving a minimum age of burial of between AD 1305 and 1525.



Figure S1. Radial plots of the single-grain D_e estimates for a) the sample collected from the side of the excavation pit; and b) the sample taken from within the cranial vault. The shaded region in each plot indicates the D_e value $\pm 2\sigma$ used to determine the burial age.

References

AITKEN, M.J. 1998. An introduction to optical dating: the dating of Quaternary sediments by the use of photon-stimulated luminescence. Oxford & New York: Oxford University Press.

BRONK RAMSEY, C. 2009. Bayesian analysis of radiocarbon dates. *Radiocarbon* 51: 337–60.

COOK, G.T., C. BONSALL, R.E.M. HEDGES, K. MCSWEENEY, V. BORONEAN & P.B. PETTITT. 2001. A freshwater diet-derived ¹⁴C reservoir effect at the Stone Age sites in the Iron Gates Gorge. *Radiocarbon* 43: 453–60.

FALLON, S.J., L.K. FIFIELD & J.M. CHAPPELL. 2010. The next chapter in radiocarbon dating at the Australian National University: status report on the single stage AMS. *Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms* 268: 898–901. http://dx.doi.org/10.1016/j.nimb.2009.10.059

FRASER, R. 2007. A study of stable carbon, nitrogen and oxygen isotopes in modern Australian marsupial herbivores, and their relationships with environmental conditions. Unpublished PhD dissertation, Australian National University.

GALBRAITH, R.F. & G.M. LASLETT. 1993. Statistical models for mixed fission track ages. *Radiation Measurements* 21: 459–70. http://dx.doi.org/10.1016/1359-0189(93)90185-c

GALBRAITH, R.F., R.G. ROBERTS, G.M. LASLETT, H. YOSHIDA & J.M. OLLEY. 1999. Optical dating of single and multiple grains of quartz from Jinmium rock shelter, northern Australia, part 1: experimental design and statistical models. *Archaeometry* 41: 339–64. http://dx.doi.org/10.1111/j.1475-4754.1999.tb00987.x

HOGG, A.G., Q. HUA, P.G. BLACKWELL, M. NIU, C.E. BUCK, T.P. GUILDERSON, T.J. HEATON, J.G. PALMER, P.J. REIMER, R.W. REIMER, C.S.M. TURNEY & S.R.H. ZIMMERMAN. 2013. SHCal13 Southern hemisphere calibration, 0–50,000 years cal BP. *Radiocarbon* 55: 1869–87. http://dx.doi.org/10.2458/azu_js_rc.55.16783

HUA, Q., M. BARBETTI & A.J. RAKOWSKI. 2013. Atmospheric radiocarbon for the period 1950–2010. *Radiocarbon* 55: 2059–72. http://dx.doi.org/10.2458/azu js rc.v55i2.16177 MEJDAHL, V. 1979. Thermoluminescence dating: beta-dose attenuation in quartz grains. *Archaeometry* 21: 61–72. http://dx.doi.org/10.1111/j.1475-4754.1979.tb00241.x

MEREDITH, K.T., S.E. HOLLINS, C.E HUGHES, D.I. CENDN, S. HANKIN & D.J.M. STONE. 2009. Temporal variation in stable isotopes (18O and 2H) and major ion concentrations within the Darling River between Bourke and Wilcannia due to variable flows, saline groundwater influx and evaporation. *Journal of Hydrology* 378: 313–24. http://dx.doi.org/10.1016/j.jhydrol.2009.09.036

MURRAY, A.S., R. MARTEN, A. JOHNSTON & P. MARTIN. 1987. Analysis for naturally occurring radionuclides at environmental concentrations by gamma spectrometry. *Journal of Radioanalytical and Nuclear Chemistry* 115: 263–88. http://dx.doi.org/10.1007/BF02037443

OLLEY, J.M., T. PIETSCH & R.G. ROBERTS. 2004. Optical dating of Holocene sediments from a variety of geomorphic setting using single grains of quartz. *Geomorphology* 60: 337–58. http://dx.doi.org/10.1016/j.geomorph.2003.09.020

PIETSCH, T.J. 2009. Optically stimulated luminescence dating of young (<500 years old) sediments: testing estimates of burial dose. *Quaternary Geochronology* 4(5): 406–22. http://dx.doi.org/10.1016/j.quageo.2009.05.013

PRESCOTT, J. R. & J.T. HUTTON. 1994. Cosmic-ray contributions to dose-rates for luminescence and ESR dating—large depths and long-term time variations. *Radiation Measurements* 23(2–3): 497–500. http://dx.doi.org/10.1016/1350-4487(94)90086-8

ROBERTS, R.G., R.F. GALBRAITH, H. YOSHIDA, G.M. LASLETT & J.M. OLLEY. 2000. Distinguishing dose populations in sediment mixtures: a test of single-grain optical dating procedures using mixtures of laboratory-dosed quartz. *Radiation Measurements* 32: 459–65. http://dx.doi.org/10.1016/S1350-4487(00)00104-9

STOKES, S., S. INGRAM, M.J. AITKEN, F. SIROCKO, R. ANDERSON & D. LEUSCHNER. 2003. Alternative chronologies for Late Quaternary (Last Interglacial–Holocene) deep sea sediment via optical dating of silt-size quartz. *Quaternary Science Reviews* 22: 925–41. http://dx.doi.org/10.1016/S0277-3791(02)00243-3 VAN KLINKEN, G.J. 1999. Bone collagen quality indicators for palaeodietary and radiocarbon measurements. *Journal of Archaeological Science* 26: 687–95. http://dx.doi.org/10.1006/jasc.1998.0385

WOOD, R.E., T.F.G. HIGHAM, A. BUZILHOVA, A. SUVOROV, J. HEINEMEIER & J. OLSEN. 2013. Freshwater radiocarbon reservoir effects at the burial ground of Minino, northwest Russia. *Radiocarbon* 55: 163–77. http://dx.doi.org/10.1017/S0033822200047883